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Stereoselective Access to Polyfunctionalized Decalins

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Abstract: As a model of a stereoselective approach to ajugareptansin 1, a versatile and straightforward access to polyfunctionalized *cis* or *trans* decalins is presented. It is based on a Lewis acid-mediated addition of furan to the unsaturated ketoester 3 followed by oxidative opening of the furan to give *cis* decalins by an aldolisation process. Selective epimerization at carbons 5 or 10 is described and allows the access to the *trans* ring geometry.

Highly polyfunctionalized decalin systems are present in many biological active natural products. Our interest in the antifeedant activities of ajugareptansin 1^2 led us to investigate new approaches aimed towards the construction of the decalinic moiety of this molecule.

In continuation with our previous work on condensation of furans³ with activated cyclohexenone in the presence of BF₃.Et₂O as catalyst, we report here the results of our efforts to control the relative configurations at carbons 3, 4, 5, 8, and 10⁴ in a model compound 2 bearing a methyl group at carbon C-8.



The Lewis acid-mediated addition of furan on the ketoester 3^5 was found to be fully controled with respect to the methyl group at C-8, leading to 4^6 in 78% yield as a single diastereomer. Oxidative ring opening of the furan system using *m*-chloroperoxybenzoic acid (*m*-CPBA) in anhydrous methanol led to a mixture of the two diastereomeric *cis* decalins 5 and 6⁷ in high yield in a 4/1 ratio. We assume that this reaction proceeds through an unsatured 1,4-dicarbonyl intermediate⁸via an intramolecular aldolisation process.³



Attempts to purify this mixture by flash chromatography on silica gel gave, in addition to compounds 5 and 6, a new product to which the structure 7⁹ was assigned as explained below. We had previously demonstrated in a similar case³ that silica gel was able to promote a retroaldolisation / aldolisation process yielding the thermodynamically more stable *trans* system.¹⁰ When the OH group at C-4 position was protected as a TBDMS ether¹¹ prior to chromatography, the *cis* decalin 8¹² was isolated in 42% overall yield. Epimerization at C-10 of the protected compound was obtained either by using DBU in anhydrous THF (63%) or quantitatively with alumina in refluxing diethyl ether.¹³ Introduction of an oxygen atom at C-3 was achieved by sodium methoxide, leading to 2¹⁵ in quantitative yield.

The structure assignments for compounds 7, 8, 9, and 2 were supported by extensive NMR experiments and the X-ray crystallography¹⁶ of compound 10 obtained quantitatively by DIBAL-H[®] reduction of compound 2.



OR **FEP** drawing of **10** (enantiomer)

For the *trans* ring junction (7, 9 and 2), H-10 resonates at 2.6, 2.6 and 2.4 ppm. For the *cis* junction (5, 6 and 8), this resonance is shifted to lower field (3.6, 3.2 and 3.3 ppm) due to the anisotropy of the close methyl ester at C-5. A high value of the coupling constant between H_a and H_c in 9 (J = 13.5 Hz) and 2 (J = 14 Hz) is in agreement with the proton H_a in axial position in both structures. In 7, H_a displays two smaller coupling constants (4 and 6 Hz), proving that H_a is in equatorial position.

The strategy developed here allows for a versatile and stereocontrolled access to either of the four highly functionalized decalins 7, 8, 9 and 2 from the easily prepared intermediate 4. These results in hand, we now have good hope to reach the *trans* decalinic moiety of ajugareptansin.

References and Notes

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- 4. Refered to the numbering of clerodans.
- Unsaturated ketoester 3 was prepared from 4,4-dimethylcyclohex-2-enone using the following sequence : 1) Mc2CuMgI, ether, 0°C (94%) ; 2) NaH, CO(OMe)2, refluxing DME (70%) ; 3) PhSeCl, pyridine, 0°C then H2O2,(82%).
- 6. All the new compounds were fully characterized by NMR, IR and MS.
- Compound 5: ¹H NMR (400 MHz, CDCl₃, ppm): 2 2.1 (m, 1H, H-8), 2.4 (dd, 1H, J = 5 and 15 Hz, H-7b), 3 (dd, 1H, J = 4.8 and 15 Hz, H-7c), 3.6 (s, 1H, H-10).
 Compound 6: ¹H NMR (400 MHz, CDCl₃, ppm): 2 2.1 (m, 1H, H-8), 2.3 (dd, 1H, J = 4 and 16 Hz, H-7b), 2.6 (dd, 1H, J = 12 and 16 Hz, H-7c), 3.2 (s, 1H, H-10).
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- Compound 7: ¹H NMR (400 MHz, CDCl₃, ppm): 2 (m, 1H, H-8), 2.18 (dd, 1H, J = 4 and 15 Hz, H-7b), 2.6 (s, 1H, H-10), 3.03 (dd, 1H, J = 6 and 15 Hz, H-7c).
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- 12. Compound 8: ¹H NMR (400 MHz, CDCl₃, ppm) : 2.1 2.2 (m, 1H, H-8), 2.3 (dd, 1H, J = 15 and 8.5 Hz, H-7b), 2.7 (dd, 1H, J = 15 and 5.5 Hz, H-7c), 3.3 (s, 1H, H-10) ; ¹³C NMR (100 MHz, CDCl₃, ppm) : 56.2 (C-10), 67.3 (C-4), 129.8 (C-2), 169.3 (C-11), 197. (C-1), 206 (C-6).
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- 14. Compound 9: ¹H NMR (400 MHz, CDCl₃, ppm): 1.6 1.7 (m, 1H, H-8), 2.3 (dd, 1H, J = 13.5 and 3.5 Hz, H-7b), 2.6 (s, 1H, H-10), 2.8 (t, 1H, J = 13.5 Hz, H-7c), 5.2 (t, 1H, J = 2 Hz, H-4), 6 (dd, 1H, J = 2 and 10.3 Hz, H-2), 6.3 (dd, 1H, J = 2.3 and 10.3 Hz, H-3); ¹³C NMR (100 MHz, CDCl₃, ppm): 60.6 (C-10), 69.4 (C-4), 130 (C-2), 144.5 (C-3), 195 (C-1), 204 (C-6).
- 15. Compound 2: ¹H NMR (400 MHz, CDCl₃, ppm) : 2.1 2.3 (m, 2H, H-2a and H-7b), 2.4 (s, 1H, H-10), 2.8 (t, 1H, J = 13.5 Hz, H-7c), 3 (dd, 1H, J = 5.5 and 14 Hz, H-2a), 3.5 3.6 (m, 1H, H-3), 4.5 (d, 1H, J = 7 Hz, H-4); ¹³C NMR (100 MHz, CDCl₃, ppm) : 52.2 and 56.3 (2 OCH₃), 61.4 (C-10), 72.7 (C-4), 81 (C-3), 172 (C-ester), 202.8 and 203.6 (C-1 and C-6).
- 16. We gratefully thank Pr. T. Prangé for X Ray crystallographic analyses.

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